

Elucidating molecular connection between IAHS onset and Alsin protein by means of Homology Modelling and Molecular Dynamics

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Abstract

The Infantile-onset Ascending Hereditary Spastic Paralysis (IAHSP) is an incurable rare neurodegenerative disease related to a mutation-driven aberrant behaviour of the Alsin protein. The lack of information on Alsin atomic structure limits a complete understanding on pathology mechanisms. In this work, molecular modelling techniques have been applied to shed lights on Alsin folding dynamics and misfunction induced by aberrant mutations.

Introduction

Alsin is a key protein involved in the onset of the IAHS, a rare pathology characterized by the degeneration of upper motor neurons of the pyramidal tract. The Alsin protein, encoded by the *Als2* gene,¹ is composed of 1657 amino acids and also expressed in the spinal cord and brain.² Alsin is predicted as formed by three structured domains, *i.e.*: i) the RCC1-like domain (RLD) in N-terminal position, ii) the central B cell lymphoma homology (DH) and pleckstrin-homology (PH) domain, and iii) the C-terminal vacuolar protein sorting 9 (VPS9) domain. *In-vitro* studies have proposed that a crucial step for the Alsin physiological pathway is the self-tetramerization and has highlighted how single point mutations in the sequence of VPS9 structured domain are correlated to a reduced tendency to oligomerise³ and thus to the onset of the pathology. Nowadays, any resolved protein model for the Alsin is available, limiting computational studies and computer-aided drug discovery investigations. For this reason, the current research will be focused on the development of a

homology model for the VPS9 domain of Alsin. The point mutation R1611W in VPS9, *i.e.* amino acid change in position 1611, Arginine to Tryptophane, altering Alsin function, will be investigated. Models developed will be employed to understand the effect of single point mutation on the stability of the structured domain and to suggest possible alteration on the self-oligomerization process.

Materials and Methods

To predict the secondary and tertiary structure, the VPS9 sequence obtained from the Uniprot database (Q96Q42) has been employed. Homology modelling has been performed through the I-Tasser and the quality of the model has been assessed with PROCHECK software. Subsequently, the obtained model of the VPS9 Wild Type (WT) has been mutated to obtain a model for the single point mutant R1611W (MUT). Three replicas of 1.5 μ s long Molecular Dynamics (MD) simulations have been carried out both for the WT and MUT systems. All systems' topology has been modelled through AMBER99sb-ildn forcefield. Systems were placed in a dodecahedron box filled with explicit water (TIP3P) and neutralized with Na⁺ and Cl⁻ ions added at a physiological concentration of 0.15 M. The engine employed for MD simulations and analysis was GROMACS 2020.3. Firstly temperature (298 K) and pressure (1 bar) have been equilibrated restraining protein positions. After removing restraints the MD production was performed at constant temperature (298 K) and pressure (1 bar). The radius of gyration (R_g) has been employed as a measure of the compactness of the structure. Results have been obtained as the time average concatenating the last 500 ns of simulations.

Results

The quality of VPS9 homology model (Figure 1A) has been evaluated through I-Tasser scores, *i.e.* the Confidence Score (C-Score) and Template Modelling Score (TM-Score) respectively equal to 1.76 and 0.96 ± 0.05 . Moreover, the stereochemistry has been validated via the Ramachandran plot, which reported 95,6 % of residues in the most favoured regions and 4,6 % in the additional allowed regions (Figure 1B).

Molecular dynamics simulation both on the WT and the MUT models have been done to understand the consequences of the mutation on the protein function. Figure 2

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reports the probability distribution of R_g for both WT and MUT VPS9 models.

Both systems show the same mean value for the distribution of R_g , but with remarkably different standard deviations (WT 15.4 ± 0.42 Å, MUT 15.1 ± 0.13 Å), showing a greater ability of the MUT system to explore a wider range of values compared to the WT.

Discussion and Conclusions

The current study represents the first step on a better comprehension of the Alsin protein both developing the first homology model for the VPS9 domain and study its dynamics via Molecular Modelling techniques. Computational results pointed attention to conformational modifications and flexibility changes in protein dynamics, induced by the R1611W mutation. Indeed, as observed *in vitro*, mutated forms of Alsin shows a reduced tendency to oligomerize. Being the R_g a measure of protein compactness, results suggest that the Wild Type (WT) can explore a broader range of conformations than the Mutated (MUT), sug-

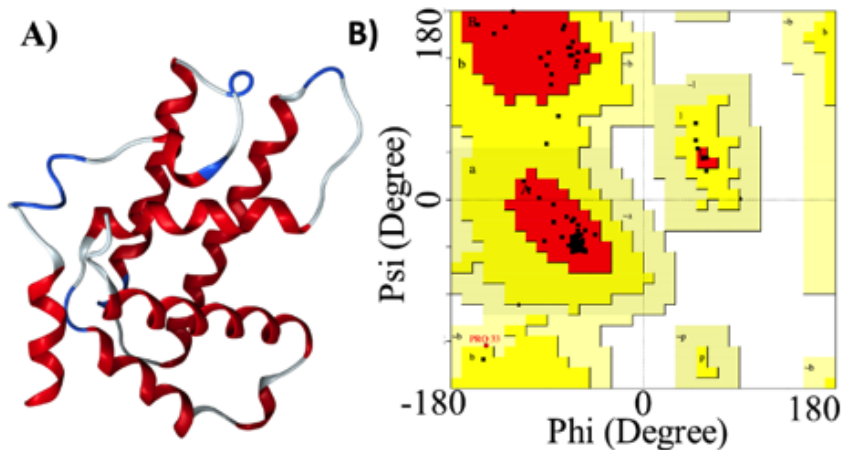


Figure 1. A) I-Tasser Homology model for the VPS9 (structured domain showed in New Cartoon representation); B) Rhamacdran Plot.

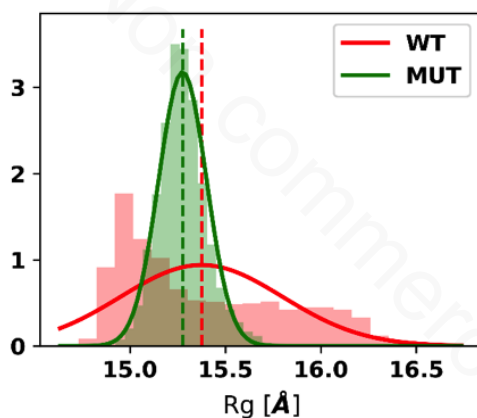


Figure 2. Probability density for Radius of Gyration for the WT (red) and the MUT (Green).

gesting for increased stiffness for the MUT form. This result can be correlated to the reduced tendency of MUT to the self-oligomerization associated to a lowered ability to rearrange its conformation and thus to be able to be engaged for protein-protein interaction and to its natural pathway. This altered behaviour can be a possible explanation on the involvement of this molecule in the onset of the IAHS. This result with the first 3D model of VPS9 developed can be a starting point for future drug screening studies aimed at restoring the physiological function of Alsln and for a better comprehension of the onset of IAHS and Alsln-related pathologies.

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