Efficacy of linezolid against Staphylococcus aureus in different rodent skin and soft tissue infections models

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Abstract

Linezolid is approved for complicated and uncomplicated skin and soft tissue infections. We have evaluated the efficacy of this drug in murine as well as in rat skin and soft tissue infection models using Staphylococcus aureus ATCC and clinical strains. In thigh infection model the dose of linezolid required for more than 1 log10 kill from baseline inoculum in neutropenic mice and rats was 100 mg/kg and 50 mg/kg, b.i.d/day, respectively, which was 5 and 4 folds more than that in immunocompetent animals, respectively. Dose required to achieve 1 log10 killing was similar against different strains of S. aureus in immunocompetent mouse thigh infection model. However, in murine groin abscess infection model, a dose of 100 mg/kg, b.i.d/day of linezolid produce static effect in 2 days, but revealed to be superior in 4 days treatment and showed approximately 1 log10, killing from base line inoculum. Based upon pharmacokinetic profile, a 24-h AUC/MIC required for linezolid efficacy in murine groin abscess model was 91.5 for the strain used in this study. As linezolid is taken as a gold standard drug in the evaluation of new chemical entity, this data could be useful for comparing the preclinical efficacy of new anti-MRSA agents.

Materials and Methods

Bacterial strains and growth conditions S. aureus L-2 and H-29 were clinical isolated and used in in vitro studies. S. aureus ATCC 25923 was purchased from the American Type Culture Collection (Manassas, Va., U.S.A). Meticillin resistant S. aureus (MRSA) 562 was obtained from Ranbaxy Research Laboratories Infectious Diseases bacterial culture collection. Isolates were grown in Trypticase soy broth (Becton Dickinson, Cockeysville, MD., U.S.A) at 37°C.

Drug preparation Linezolid was procured from the National Chemical Laboratory (Pune, India). Vancomycin was obtained from API manufacturing facility of Ranbaxy Research Laboratories. Levofloxacin was purchased from commercial sources (Levaquin tablets, ORTHO-MCNEIL Pharmaceutical Inc. Raritan, NJ). Linezolid was prepared in 0.25% methyl cellulose. Vancomycin and levofloxacin were dissolved in sterile milli-Q water.

Susceptibility testing MICs were determined by the microbroth dilution method according to CLSI guidelines.6 The MIC was defined as the lowest concentration of antibiotic that prevented visible growth after an incubation period of 18 to 24 h.7

Experimental animals Swiss albino mice (5-6 weeks old, 18-22 g weight and specific-pathogen free) and Wistar rats (4-6 weeks old, 90-110 g weight, and specific-pathogen free) were used for the studies. The experimental protocols were approved by the Institutional Animal ethics Committee (IAEC) of Ranbaxy Research Laboratories, Gurgaon, India. Animals were adapted to standardized environmental conditions for 2-3 days before the experiments. Thigh infection was performed in immunocompetent and neutropenic animals. In each group 6 animals were taken. Animals were rendered neutropenic (neutrophils <100/mm3) as described earlier by Andes et al.,8 by injecting cyclophosphamide (Lodoxan, Darbaur Pharmaceuticals, New Delhi, India) at 150 mg/kg (day − 4) and 100 mg/kg (day − 1) and 100 mg/kg (day − 4) and 80 mg/kg (day − 1) in mice and rat, respectively. For groin infection in mice the hairs were trimmed from the groin area of the animals using an electric hair clipper (TSE systems, Germany) and subsequently hair remover cream (Wyeth, USA) was applied to completely remove the hairs, a day before infection.

References

1. Community acquired methicillin resistant S. aureus (CA-MRSA) not only causes the skin and soft tissue infection but is also responsible for highly invasive and rapidly progressive, life-threatening diseases such as necrotizing pneumonia, severe sepsis and necrotizing fasciitis.2

2. Linezolid, the first marketed oxazolidinone antibiotic, has broad spectrum in vitro activity against antibiotic-susceptible and resistant Gram-positive bacteria, including MRSA and S. aureus with intermediate resistance to glycopeptides.3 Plasma concentrations of intravenous and oral linezolid are equivalent, with average concentrations exceeding the MICs for susceptible pathogens throughout the 12 h dosing interval.4 The drug is indicated for complicated and uncomplicated skin infection caused mainly by S. aureus.5 However, the efficacy of this drug is not much assessed in animal model of skin and soft tissue infection or available data is fragmented in nature.

3. The objective of the present study is to evaluate and compare the efficacy of linezolid against various S. aureus strains in an immunocompetent and neutropenic mouse as well as rat thigh infection models and in murine groin abscess model, which is a more complicated skin and soft tissue infection model (cSSTI). Furthermore, the efficacy data is supported with the relevant pharmacokinetic data for both the species.

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Pharmacokinetic study in mouse and rat

To correlate the doses of linezolid administered to mice and rat with measures of exposure, single-dose pharmacokinetic studies were conducted with Swiss albino mice and Wister rats. Twenty-seven male Swiss albino were taken for the study. A dose of 5 mg/kg and 100 mg/kg of linezolid was orally administered to three mice in each group. Terminal blood samples (collected into labeled tubes containing 25 μL of 20% w/v sodium citrate) were taken from animals at 15 min, 30 min, 60 min, 2, 4, 8, 12, 24 and 48 h. Plasma was separated from blood samples and stored at -70°C until analysis. Pharmacokinetics and oral bioavailability in male Wistar rats was studied at 50 mg/kg oral. Blood was collected from three animals at 15 min, 30 min, 60 min, 2, 4, 8, 12 and 24 h. Plasma was separated from blood samples and stored at -70°C as described above. The level of linezolid in plasma was determined by LC-MS/MS. The lower limit of quantitation (LLOQ) was 14.84 ng/mL.

Localized thigh infection (immunocompetent versus neutropenic)

Thigh infection was established in both immunocompetent and neutropenic mice and rat. For infection, overnight grown culture in BHI (brain heart infusion) was adjusted to contain approximately 3×10^6 CFU/mL of S. aureus. The culture was diluted 1:10 in normal saline (NS) and mixed with 5% hog gastric mucin in a 1:1 proportion ratio to achieve an inoculum of approximately 1.5×10^8 CFU/mL. For establishing infection, 100 μL and 250 μL of this culture was injected intramuscularly into the right thigh of immunocompetent mice and rat, respectively.

For neutropenic animals, the initial inoculum was diluted 1:100 in NS and then mixed with 5% hog gastric mucin in 1:1 ratio to get an inoculum of approximately 1.5×10^6 CFU/mL. The treatment was started 1 h post infection and administered twice daily by oral route for either 2 or 4 days. The efficacy of linezolid was evaluated at several dose levels. In order to determine the initial bacterial load, an early infection control (1 h) was dissected at the start of treatment. At the end of treatment, all groups of animals were sacrificed by cervical dislocation. The thigh muscles were dissected out under aseptic conditions and homogenized in 1 mL of NS in case of mice thigh and 5 mL NS in case of rat thigh. The bacterial count was quantified by plating the serial dilutions of homogenate on TSA and incubating at 37°C for overnight.

Groin abscess model

For causing groin abscess infection, advent semi solid nutrient agar was prepared by suspending 0.6% Bactoagar (Becton Dickinson and Company, Sparks MD, USA) in nutrient broth (HiMedia Laboratories, Mumbai, India) and sterilized by autoclaving. The overnight grown culture in BHI was diluted 1:20 in normal saline and then 1:10 in 0.6% semi solid nutrient agar. The animals were injected subcutaneously with 0.5 mL of the prepared inoculum containing approximately 10^6 CFU of bacteria in the groin area.

Mice received linezolid by oral and vancomycin by subcutaneous route, b.i.d. for either 2 or 4 days. After 18 h of the last dose, animals were euthanized; the skin was disinfected with 70% alcohol and allowed to dry. The abscesses formed in the groin region were aseptically removed along with the skin and homogenized in 1 mL of sterile NS. Serial ten-fold dilutions of the homogenates were plated on TSA and the viable bacterial numbers were determined. Experiment was performed in duplicate and results combined for analysis.

Data analysis

The in vivo efficacy data was analysed using GraphPad Prism version 4.1 and Microsoft Excel 2007. Pharmacokinetic data was analysed with the WinNonlin software (Pharsight, Mountain View, CA, USA) program version 4.1. Linezolid efficacy data was expressed in the form of change in bacterial load per thigh from base line inoculum (bacterial load at the start of treatment).

Results

Susceptibility profile of S. aureus strains and exposure study

The minimum inhibitory concentration of S. aureus strains used in this study is presented in Table 1. The systemic plasma exposure profile of linezolid in Swiss albino mice and Wister rats following different oral doses is given in Table 2 and Table 3 respectively. The oral exposure study of linezolid in mice was performed at 5 and 100 mg/kg, however, PK values of 20, 40 and 80 mg/kg were extrapolated. Linezolid oral exposure study in rats was performed at 50 mg/kg and the values of 10 and 100 mg/kg were extrapolated.

S. aureus thigh infection

Four strains of S. aureus (ATCC 25923, MRSA H-29, MRSA L-2 and MRSA-562) were used for efficacy evaluation of linezolid in murine thigh infection model. The oral efficacy of linezolid was evaluated at a dose of 20 mg/kg, bid in 1 and 2 days murine SSTI models and summarized in Table 1. In murine SSTI model, all tested S. aureus strains showed good infectivity and grew from 6.3±0.25 to 8.6±0.42

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Table 1. Mean log_{10} reduction in cfu/thigh of S. aureus by linezolid in thigh and groin infection studies.

<table>
<thead>
<tr>
<th>Strains</th>
<th>MIC (μg/mL)</th>
<th>Mean log_{10} reduction in CFU/thigh</th>
<th>Mean log_{10} reduction in CFU/abscesses</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Day 1 (Mean±SD)</td>
<td>Day 2 (Mean±SD)</td>
</tr>
<tr>
<td>S. aureus</td>
<td></td>
<td>Linezolid 20 mpk, BID, PO</td>
<td>Linezolid 100 mpk, BID, PO</td>
</tr>
<tr>
<td>ATCC 25923</td>
<td>1</td>
<td>3.35±0.2</td>
<td>3.92±0.62</td>
</tr>
<tr>
<td>MRSA 562</td>
<td>2</td>
<td>3.23±0.22</td>
<td>3.41±0.34</td>
</tr>
</tbody>
</table>

Table 2. Selected pharmacokinetics parameter of linezolid estimated in Swiss albino mice.

<table>
<thead>
<tr>
<th>Dose (mg/kg)</th>
<th>C_{max} (μg/mL)</th>
<th>AUC (μg.h/mL)</th>
<th>AUC/MIC ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>2.53</td>
<td>5.32</td>
<td>2.66</td>
</tr>
<tr>
<td>20*</td>
<td>16.90</td>
<td>36.50</td>
<td>18.25</td>
</tr>
<tr>
<td>40*</td>
<td>33.72</td>
<td>73.04</td>
<td>36.52</td>
</tr>
<tr>
<td>100*</td>
<td>84.35</td>
<td>182.79</td>
<td>91.40</td>
</tr>
</tbody>
</table>

*Extrapolated values.

Table 3. Selected pharmacokinetics parameter of linezolid estimated in Wister rats.

<table>
<thead>
<tr>
<th>Dose (mg/kg)</th>
<th>C_{max} (μg/mL)</th>
<th>AUC (μg.h/mL)</th>
<th>AUC/MIC ratio</th>
</tr>
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<tbody>
<tr>
<td>10</td>
<td>3.00</td>
<td>28.00</td>
<td>14.00</td>
</tr>
<tr>
<td>50</td>
<td>17.70</td>
<td>107.00</td>
<td>53.50</td>
</tr>
<tr>
<td>100*</td>
<td>29.90</td>
<td>277.00</td>
<td>138.50</td>
</tr>
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</table>
log₁₀ CFU/thigh in day 1 and to 9.0±1.3 log₁₀ CFU/thigh in 2 days. The bacterial load reduction of linezolid at a dose of 20 mg/kg in thigh infection model against all tested strains is given in the Table 1. Other standard drugs vancomycin and levofloxacin (data not presented) showed more than 4 log₁₀ reduction from late untreated control in both studies.

Efficacy in immunocompetent versus neutropenic animals

In order to access the Staphylococcal killing potential of linezolid in animal SSTI model, a detail efficacy study against S. aureus ATCC 25923 was performed in immunocompetent and neutropenic mice and rats thigh infection model. In immunocompetent infection model in mice with sensitive strain, S. aureus ATCC 25923, linezolid exhibited more than 3 log₁₀ reduction compared to the late untreated controls (> 1 log₁₀ killing from initial inoculum) at the dose of 20 mg/kg BW, bid/day. However, in murine neutropenic thigh infection model the drug showed the similar effect only at a higher dose of 100 mg/kg BW, bid/day, stasis at 80 mg/kg BW, bid/day (Figure 1). A similar efficacy of linezolid was observed in Wistar rats. A dose of 10 mg/kg BW bid/day showed >1 log₁₀ killing from initial CFU in immunocompetent rats, whereas in neutropenic rats a dose of 50 mg/kg BW, b.i.d/day is required to get similar efficacy (Figure 2).

S. aureus groin abscess model

The efficacy of linezolid was determined in groin infection model with virulent MRSA strains, L-2 and H-29 (Table 1). Using 0.6% agar as an adjuvant, the efficacy of linezolid was evaluated at several doses (Figure 3). A dose of 100 mg/kg b.i.d showed static effect in 2 days model, while in 4 days treatment approximately 1 log₁₀ killing from base line inoculum was achieved at the same dose (Figure 3).

Discussion

Complicated skin and soft tissue infections (cSSTi) caused by S. aureus are frequently encountered in clinical practice and are the significant cause of morbidity and mortality in hospitalized patients. Linezolid is the first oxazolidinone that inhibits the bacterial protein synthesis. It possesses a broad spectrum of in vitro and in vivo activity against both community and nosocomial Gram-positive pathogens. Of the many indications for which the drug has been approved, SSTI and cSSTI caused by MSSA and MRSA strains are the main area where the drug has demonstrated clinical efficacy.5,9
In the preclinical lead optimization or drug discovery, linezolid is taken as a gold standard for comparing novel compounds against *S. aureus* especially in SSTI and cSSTI models. But limited preclinical data is available on the *in vivo* efficacy of linezolid.\(^\text{10,11}\) Further, the published data of linezolid on the animal models of SSTI against *S. aureus* is scarce.

Pucci et al.,\(^\text{11}\) studied the preclinical efficacy of linezolid at 20 mg/kg in neutropenic murine thigh infection and failed to achieve any bacterial log\(_{10}\) reduction from initial control. In the present study the efficacy of linezolid was tested at various concentrations against different MSSA and MRSA including the L strain in immunocompetent and neutropenic murine thigh infection. Linezolid showed more than 1 log\(_{10}\) killing from initial-control in immunocompetent mice and more than 3 log\(_{10}\) from late infection control at 20 mg/kg, PO, b.i.d. X 1 day. Based on the AUC obtained in our exposure study, a dose of 20 mg/kg, BW bid/day gives a 24-h AUC/MIC 18.23. This is a minimum necessary dose for linezolid efficacy against *S. aureus* in cSSTI in immune-competent mice, while a 24-h AUC/MIC ratio required to produce a static effect in neutropenic mice against *S. aureus* (91.4) was 5-fold greater than those observed in immunocompetent mice. A dose of 80 mg/kg was required to achieve stasis and 1 log\(_{10}\) killing from initial-control was achieved with a dose of 100 mg/kg in neutropenic mice. This observation was also correlated well with rat thigh infection model, in which a 24-h AUC/MIC ratio required to achieve approximately 1 log\(_{10}\) reduction from initial control in neutropenic rat was 4 times higher than that required in immunocompetent rat against *S. aureus* (Table 3). Our results are in line with that reported by Andes et al.,\(^\text{8}\) a 24-h AUC/MIC as the key parameter which determines efficacy in neutropenic mice and the ratio of 90 - 100 is required for the *S. aureus* strains with an *in vitro* MIC of 1-2 \(\mu\)g/mL.\(^\text{4}\) Also this AUC best correlates with the levels achieved in humans after the recommended 600 mg b.i.d. dose for SSTI infections.\(^\text{2}\) Moreover, we have showed the detailed efficacy study of linezolid against different clinical isolates of *S. aureus* and also dose response in immunocompetent and neutropenic mice and rats SSTI models.

According to a previous report, prevention of abscess formation by antibiotic treatment based upon the minimal inhibitory concentra-

### References