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## **ROGERS MJ**

Bone Research Group, Institute of Medical Sciences, University of Aberdeen, Aberdeen, Scotland, UK

## New insights into the molecular actions of nitrogen-containing bisphosphonates

bout 10 years ago it was found that mammalian cells could convert some bisphosphonates (BPs) intracellularly into methylene-containing (AppCp-type) analogues of ATP, that accumulate in the cytosol,<sup>1,2</sup> reviewed recently.<sup>3</sup> Only simple BPs (ie those that most closely resemble PPi, such as clodronate and etidronate, but not alendronate, pamidronate or other N-BPs) appear to be metabolised.<sup>2</sup> More recently, we have confirmed that osteoclasts in vivo are capable of metabolising clodronate to AppCCl2p,<sup>4</sup> and shown that the accumulation of the AppCCl2p metabolite of clodronate in osteoclasts in vitro inhibits bone resorption by inducing osteoclast apoptosis,<sup>4</sup> perhaps by inhibiting the adenine nucleotide translocase (ANT), a component of the mitochondrial permeability transition pore.<sup>5</sup> Nitrogen-containing bisphosphonates (N-BPs), which are several orders of magnitude more potent at inhibiting bone resorption in vivo than the simple BPs,<sup>6,7</sup> are not metabolised<sup>2</sup> but are potent inhibitors of farnesyl diphosphate synthase (FPP synthase), a key enzyme of the mevalonate pathway. This enzyme is inhibited by nanomolar concentrations of N-BPs.<sup>8-10</sup> Zoledronic acid, and the structurally similar minodronate, are extremely potent inhibitors of FPP synthase<sup>10</sup> and inhibit enzyme activity even at picomolar concentrations. Importantly, studies with recombinant human FPP synthase revealed that minor modifications to the structure and conformation of the R<sup>2</sup> side chain that were known to affect anti-resorptive potency on osteoclasts<sup>10</sup> or activation of  $\gamma$ . $\delta$ -T cells<sup>11</sup> also affect the ability to inhibit FPP synthase (reviewed recently;<sup>6</sup> thus finally explain the relationship between BP structure and anti-resorptive potency. Furthermore, these studies strongly suggested that FPP synthase is the major pharmacologic target of N-BPs in osteoclasts and other cell types.

The exact mechanism by which N-BPs inhibit FPP synthase is only just becoming clear. The recent generation of an x-ray

crystal structure of the human FPP synthase enzyme, co-crystallised with risedronate or zoledronic acid, 12,13 indicates that the BPs appear to bind in one of the two isoprenoid lipid binding pockets in the enzyme active site (that would normally bind GPP or DMAPP), with the R<sup>2</sup> side chain positioned in the hydrophobic cleft that normally accomodates an isoprenopid lipid, and the phosphonate groups bound to a cluster of 3 magnesium ions. Interestingly, the nitrogen atom in the  $R^2$  side chain appears to form hydrogen bonds with a critical, conserved threonine residue. This is consistent with the earlier suggestion by Oldfield and colleagues that N-BPs mimick the structure of the enzyme's natural isoprenoid pyrophosphate substrates GPP/DMAPP and act as carbocation transition state analogues or reactive intermediates.<sup>14</sup> competing for binding at the GPP/DMAPP substrate binding pocket in the active site of the enzyme. N-BPs also appear to inhibit bacterial FPP synthase in a similar manner.<sup>15</sup>

Enzyme kinetic analysis with human FPP synthase indicates that the interaction with N-BPs is highly complex and characteristic of slow-tight binding inhibition. Initially, N-BPs appear to compete directly with DMAPP or GPP for binding to the DMAPP/GPP binding pocket. However, this is followed by more complex interactions that affect the binding of IPP. Recent computer modelling studies suggested that a second molecule of N-BP might also bind to the (IPP) isoprenoid lipid pocket in the enzyme, which appeared to be supported by enzyme kinetic analysis showing biphasic modes of inhibition.<sup>16</sup> However, binding of N-BP to the IPP pocket of FPP synthase has not yet been confirmed in x-ray crystal structures, 12,13 and the kinetic studies suggesting inhibition at the IPP site may reflect more complex conformational changes in the structure of the enzyme due to binding of N-BP in the DMAPP/GPP pocket. Nevertheless, these studies are beginning to key insights into the reasons why minor changes to the structure of the N-BP side chain or to the phosphonate groups markedly influence the potency of N-BPs for inhibiting FPP synthase, hence explaining at the detailed molecular level the structure-activity relationships of N-BPs for inhibiting bone resorption and for activating  $\gamma$ , $\delta$ -T cells, which are both due to inhibition of FPP synthase.<sup>6,11,17</sup>

## References

- Frith JC, MonkkonenJ, Blackburn GM, Russell RG, Rogers MJ. Clodronate and liposome-encapsulated clodronate are metabolized to a toxic ATP analog, adenosine 5'-(beta, gamma- dichloromethylene) triphosphate, by mammalian cells *in vitro*. J Bone Miner Res 1997;12:1358-67.
- Benford HL, Frith JC, Auriola S, Monkkonen J, Rogers MJ. Farnesol and geranylgeraniol prevent activation of caspases by aminobisphosphonates: biochemical evidence for two distinct pharmacological classes of bisphosphonate Drugs Mol Pharmacol 1999; 56:131-40.
- Rogers MJ. From molds and macrophages to mevalonate: a decade of progress in understanding the molecular mode of action of bisphosphonates. Calcif Tissue Int 2004;75:451–1.
- 4. Frith JC, Monkkonen J, Auriola S, Monkkonen H, Rogers MJ. The molecular mechanism of action of the anti-resorptive and antiinflammatory drug clodronate: evidence for the formation *in vivo* of a metabolite that inhibits bone resorption and causes osteoclast and macrophage apoptosis. Arth Rheum 2001;44:2201–10.
- Lehenkari PP, Kellinsalmi M, Napankangas JP, Ylitalo KV, Monkkonen J, Rogers MJ. Further insight into mechanism of action of clodronate: inhibition of mitochondrial ADP/ATP translocase by a nonhydrolyzable, adenine-containing metabolite. Mol Pharmacol 2002;61:1255-62.
- 6. Rogers MJ. New insights into the molecular mechanisms of action of bisphosphonates. Curr Pharm Des 2003;9:2643-58.
- Green JR, Rogers MJ. Pharmacologic profile of zoledronic acid: a highly potent inhibitor of bone resorption. Drug Dev Res 2002; 55;210-24.
- 8. van Beek E, Pieterman E, Cohen L, Lowik C, Papapoulos S. Farne-

syl pyrophosphate synthase is the molecular target of nitrogencontaining bisphosphonates. Biochem Biophys Res Commun 1999;264:108-11.

- 9. Bergstrom JD, Bostedor RG, Masarachia PJ, Reszka AA, Rodan G. Alendronate is a specific, nanomolar inhibitor of farnesyl diphosphate synthase. Arch Biochem Biophys 2000;373:231-41.
- Dunford JE, Thompson K, Coxon FP, Luckman SP, Hahn FM, Poulter CD, Ebetino F H, Rogers MJ. Structure-activity relationships for inhibition of farnesyl diphosphate synthase *in vitro* and inhibition of bone resorption *in vivo* by nitrogen-containing bisphosphonates. J Pharmacol Exp Ther 2001;296:235-42.
- Thompson K, Rogers MJ. Statins prevent bisphosphonate-induced gamma,delta-T-cell proliferation and activation *in vitro*. J Bone Miner Res 2004;19:278–88.
- Rondeau JM, Bitsch F, Geiser M, Hemmig R, Kroemer, Lehmann S. Structural basis for the exceptional *in vivo* efficacy of bisphosphonate drugs. J Med Chem in press.
- Kavanagh K, Guo K, Wu X, Knapp S, EbetinoFH, Dunford JE. Human farnesyl diphosphate synthase (FDPS): crystal structure and molecular interactions with nitrogen-containing bisphosphonates. J Bone Miner Res 2005;20:95-6.
- Martin MB, Arnold W, Heath HT, Urbina JA, Oldfield E. Nitrogen-Containing Bisphosphonates as Carbocation Transition State Analogs for Isoprenoid Biosynthesis. Biochem Biophys Res Commun 1999;263:754-8.
- Hosfield DJ, Zhang Y, Dougan DR, Brooun A, Tari LW, Swanson RV. Structural basis for bisphosphonate-mediated inhibition of isoprenoid biosynthesis. J Biol Chem 2003;279:8526–9.
- Ebetino FH, Roze CN, McKenna CE, Barnett BL, Dunford JE, Russell RGG, et al. Molecular interactions of nitrogen-containing bisphosphonates within farnesyl diphosphate synthase. J Organomet Chem 2005;690:2679-87.
- Gober HJ, Kistowska M, Angman L, Jeno P, Mori L, De Libero G. Human T cell receptor gammadelta cells recognize endogenous mevalonate metabolites in tumor cells. J Exp Med 2003;197:163– 8.