Anti-tumor activity of CCI-779 in relapsed mantle cell lymphoma

[haematologica reports] 2005;1(8):92-94

ANSELL SM GEYER SM KURTIN PJ ROWLAND KM FLYNN PJ MORTON RF DAKHIL SR GROSS HM MAURER MJ KAUFMANN SH WITZIG TE

Mayo Clinic, Rochester, MN 55905, USA

Mantle cell lymphoma (MCL) repre-
sents approximately 8% of cases of
non-Hodgkin's lymphoma (NHL)
and is an incurable aggressive B-cell maligsents approximately 8% of cases of non-Hodgkin's lymphoma (NHL) and is an incurable, aggressive B-cell malignancy. The disease usually presents in an advanced stage and commonly involves extranodal sites such as the gut, bone marrow, and peripheral blood. Most patients are older adults and there is a male predominance. The characteristic tumor cell immunophenotype is a population of CD20+ , CD10- , CD5+ , CD23- B-cells with monoclonal light chain expression on the cell surface.

MCL is also unique in that the tumor cells have a $t(11;14)(q13;q32)$ chromosomal translocation that juxtaposes the cyclin D1 gene on chromosome 11 to the immunoglobulin heavy chain enhancer region on chromosome 14.1-3 The transcription enhancers on 14q32, now linked to the cyclin D1 gene, result in the characteristic overexpression of cyclin D1 in the MCL tumor cells. There is currently no standard therapy for newly diagnosed or relapsed MCL. Many regimens have shown significant activity in this disease, $4-17$ but relapse typically occurs, and patients usually die of progressive disease, with a median survival of 3 to 4 years. It is therefore clear that new treatments are needed for MCL.

Targeting molecular pathways

Even though cyclin D1 mRNA is constitutively expressed in MCL, it is subject to translational regulation by a pathway involving the mammalian target of rapamycin (mTOR).18,19 Activated receptor tyrosine kinases and activated ras proteins enhance the catalytic activity of the lipid kinase phosphatidylinositol-3 kinase (PI3K), which converts phosphatidylinositol-4,5 bisphosphate (PIP2) to phosphatidylinositol-3,4,5-trisphosphoate (PIP3). PIP3 activates the protein kinase phosphoinositidedependent kinase 1 (PDK1), which, along with a second kinase such as integrin-linked kinase (ILK), contributes two phosphorylations required for maximal Akt activity. Akt then phosphorylates a number of substrates,

including tuberous sclerosis (TSC) protein 2 (TSC2), which in its unphosphorylated state is complexed with TSC protein 1 (TSC1) and acts as a GTPase activating protein that diminishes activation of the small guanine nucleotide binding protein Rheb. When the TSC1/TSC2 complex is inactivated by Akt, Rheb remains in a GTPbound state that activates mTOR, a protein kinase that regulates mRNA translation by phosphorylating two critical substrates, eukaryotic initiation factor (eIF) 4E (eIF4E) binding protein (4E-BP1) and p70S6 kinase.^{20,21}

Previous studies have shown that eIF4E is a component of a helicase complex that binds to the cap structure at the 5# end of mRNAs and enhances the ability of ribosome-eIF complexes to scan the mRNA in search of a translation initiation site.²² The ability of eIF4E to bind to and participate in this helicase complex is inhibited when 4E-BP1 is bound. This inhibitory interaction is possible only when 4E-BP1 is unphosphorylated and is abrogated when 4E-BP1 is sequentially phosphorylated by mTOR and other kinases.^{22,23} At the same time, mTOR mediated phosphorylation activates p70S6K, enabling its phosphorylation of ribosomal protein S6 and possibly other substrates, thereby enhancing the translation of messages with 5# terminal oligopyrimidine tracts.18,22 Collectively, these events markedly enhance translation of a small but important group of messages, including those encoding c-myc, ornthithine decarboxylase, and cyclin D1, as well as ribosomal proteins themselves.18,22,24,25 mTOR activity is modulated by mitogenic signals, which are transmitted through a signal transduction pathway involving PI3K, Akt, and TSC1 and TSC2.18,19,26,27 In addition, mTOR-mediated signaling is also subject to modulation by the macrocyclic lactone rapamycin and its derivatives.19,26,27 Once these agents bind to the 12 kDa cytosolic FK506-binding protein FKBP12, the resulting rapamycin-FKBP12 complexes bind to a specific site near the catalytic domain of mTOR and inhibit phosphorylation of mTOR substrates by a mechanism that remains somewhat poorly understood.²⁷

As a consequence, translation of messages that require mTOR signaling is inhibited. This mechanism is thought to be responsible for the immunosuppressive effects of rapamycin as well as its putative antineoplastic activity.

CCI-779 (also known as temsirolimus), a dihydroester of rapamycin that is suitable for intravenous use, is currently undergoing testing in solid tumor patients as a potential antineoplastic agent.28-31 In view of the role of cyclin D1 in MCL, we conducted an initial phase II trial of single-agent temsirolimus at a dose level of 250 mg IV weekly for patients with relapsed MCL.³² This study with weekly CCI-779 at a dose of 250mg demonstrated encouraging antitumor activity but thrombocytopenia was frequently observed and was dose limiting. Based on results from clinical trials in other malignancies, a subsequent cohort of patients was enrolled on the study and received a low-dose (25 mg) of CCI-779 to determine if a 10-fold decrease in the dose could produce a similar overall response rate.³³

Clinical results

In the initial patient cohort in the study32, patients with relapsed or refractory MCL were eligible to receive temsirolimus 250 mg intravenously every week as a single agent. In the subsequent cohort of patients enrolled in the study33, patients received CCI-779 25mg IV weekly as a single agent. Eligible patients had biopsy proven cyclin D1 positive MCL and had relapsed or were refractory to therapy. Patients were required to have measurable disease, an adequate performance status, and adequate organ and bone marrow reserve. Patients were restaged after 1 cycle (4 doses), after 3 cycles, and every 3 cycles thereafter. Patients with a tumor response after six cycles were eligible to continue drug for a total of 12 cycles or two cycles after complete remission (CR), and were then observed without maintenance treatment.

In the initial study cohort, using temsirolimus at the 250mg dose level, 35 patients were enrolled and were assessable for toxicity. One patient had MCL by histology but was cyclin D1 negative and was ineligible for efficacy. The median age was 70 years (range, 38 to 89 years), 91% were stage 4, and 69% had two or more extranodal sites. Patients had received a median of three prior therapies (range, one to 11), and 54% were refractory to the last treatment. The overall response rate was 38% (13 of 34 patients; 90% CI, 24% to 54%) with one complete response (3%) and 12 partial responses (35%). The median time-to-progression in all patients was 6.5 months (95% CI, 2.9 to 8.3 months), and the duration of response for the 13 responders was 6.9 months (95% CI, 5.2 to 12.4 months). Hematologic toxicities were the most common, with 71% (25 of 35 patients) having grade 3 and 11% (four of 35 patients) having grade 4 toxicities. Thrombocytopenia was the most frequent cause of dose reductions but was of short duration, typically resolving within 1 week. However, dose reductions were necessary in the majority of patients.

Due to the finding that thrombocytopenia was the dose limiting factor, and the fact that dose intensity of the drug did not appear to correlate with clinical response, we decided to explore the use of a lower dose. Responses had been seen in other malignancies using 25mg IV weekly, and a second cohort of patients were enrolled and received a substantially lower dose of 25mg weekly IV. In this subsequent study cohort, using the 25mg dose level, 26 of the planned 27 patients have been enrolled. The design-mandated interim analysis on the first 13 evaluable patients confirmed 7 responders (54%; CI 16%-84%) and stable disease in the remaining 6 patients. One patient (8%) had a complete response and 6 (46%) had a partial response. Accrual to this study is almost complete and a final analysis will then be done. Significantly less thrombocytopenia was seen with the lower dose, but the frequency of anemia and neutropenia was similar in both studies.

In both studies, the most common adverse events of all grades were thrombocytopenia, hyperglycemia, anemia, neutropenia, increased triglycerides, mucositis, fatigue, infection without concomitant neutropenia, rash, nausea, weight loss, AST elevations, abnormal sense of taste, loss of appetite, hypercholesterolemia, and sensory neuropathy. No grade 5 events due to treatment were reported.

Pharmacodynamics

To develop an assay for mTOR inhibition that could be applied in clinical MCL samples, we initially treated MO258 cells with varying concentrations of rapamycin *in vitro* and probed whole cell lysates for phosphorylation of substrates downstream of mTOR using commercially available antiphosphoepitope antibodies. Blotting for phospho–4E-BP1 in this cell line and others proved difficult. In contrast, we observed that phospho-S6 and, with somewhat more difficulty, phosphop70S6 kinase could be demonstrated. Moreover, inhibition of S6 phosphorylation was readily detectable at 0.1 nmol/L rapamycin and essentially complete at 1 nmol/L.

When this assay was applied to MCL samples from the blood of patients receiving temsirolimus 250mg weekly in the initial cohort, phosphorylation of S6 was more readily detectable than phosphorylation of p70S6K. Examination of serial samples revealed two distinct patterns. First, S6 phosphorylation was inhibited after temsirolimus treatment in three patients. Of these three patients, one responded to therapy, one was stable, and one progressed without ever responding. In contrast, there was no evidence that S6 phosphorylation was inhibited in circulating MCL cells from two other patients. One of these patients had a PR; the other progressed on therapy.

Conclusions

Single-agent temsirolimus has substantial antitumor

References

- 1. Bertoni F, Zucca E, Cotter FE: Molecular basis of mantle cell lymphoma. Br J Haematol 124:130-140, 2004
- 2. Kurtin PJ: Mantle cell lymphoma. Adv Anat Pathol 5:376-398, 1998
- 3. Kurtin PJ, Hobday KS, Ziesmer S, et al: Demonstration of distinct antigenic profiles of small B-cell lymphomas by paraffin section immunohistochemistry. Am J Clin Pathol 112: 319-329, 1999
- 4. Blay JY, Sebban C, Surbiguet C, et al: High-dose chemotherapy with hematopoietic stem cell transplantation in patients with mantle cell or diffuse centrocytic non-Hodgkin's lymphomas: A single center experience on 18 patients. Bone Marrow Transplant 21:51-54, 1998
- 5. Bosch F, Lopez-Guillermo A, Campo E, et al: Mantle cell lymphoma: Presenting features, response to therapy, and prognostic factors. Cancer 82:567-575, 1998
- 6. Coiffier B, Haioun C, Ketterer N, et al: Rituximab (anti-CD20 monoclonal antibody) for the treatment of patients with relapsing or refractory aggressive lymphoma: A multicenter phase II study. Blood 92:1927-1932, 1998
- Decaudin D, Bosq J, Munck JN, et al: Mantle cell lymphomas: Characteristics, natural history and prognostic factors of 45 cases. Leuk Lymphoma 26:539- 550, 1997
- 8. Decaudin D, Bosq J, Tertian G, et al: Phase II trial of fludarabine monophosphate in patients with mantle-cell lymphomas. J Clin Oncol 16: 579-583, 1998
- 9. Dreger P, von Neuhoff N, Kuse R, et al: Sequential high-dose therapy and autologous stem cell transplantation for treatment of mantle cell lymphoma. Ann Oncol 8:401-403, 1997
- Foran JM, Cunningham D, Coiffier B, et al: Treatment of mantle-cell lymphoma with Rituximab (chimeric monoclonal anti-CD20 antibody): Analysis of factors associated with response. Ann Oncol 11:117-121, 2000
- 11. Foran JM, Rohatiner AZ, Cunningham D, et al: European phase II study of rituximab (chimeric anti-CD20 monoclonal antibody) for patients with newly diag-

nosed mantle-cell lymphoma and previously treated mantle-cell lymphoma, immunocytoma, and small B-cell lymphocytic lymphoma. J Clin Oncol 18: 317-24, 2000

- 12. Freedman AS, Neuberg D, Gribben JG, et al: High-dose chemoradiotherapy and anti-B-cell monoclonal antibodypurged autologous bone marrow transplantation in mantle-cell lymphoma: No evidence for long-term remission. J Clin Oncol 16:13-18, 1998
- 13. Ghielmini M, Schmitz SF, Burki K, et al: The effect of Rituximab on patients with follicular and mantle-cell lymphoma: Swiss Group for Clinical Cancer Research (SAKK). Ann Oncol 11:123- 126, 2000
- 14. Gopal AK, Rajendran JG, Petersdorf SH, et al: High-dose chemo-radioimmunotherapy with autologous stem cell support for relapsed mantle cell lymphoma. Blood 99:3158-3162, 2002
- 15. Inwards D, Brown D, Fonseca R, et al: NCCTG phase II trial of 2-chlorodeoxyadenosine (2-CDA) as initial therapy for mantle cell lymphoma: A welltolerated treatment with promising activity. Blood 94:660a, 1999 (suppl 1)
- 16. Howard OM, Gribben JG, Neuberg DS, et al: Rituximab and CHOP induction therapy for newly diagnosed mantlecell lymphoma: Molecular complete responses are not predictive of progression-free survival. J Clin Oncol 20:1288- 1294, 2002
- 17. Khouri IF, Romaguera J, Kantarjian H, et al: Hyper-CVAD and high-dose methotrexate/cytarabine followed by stemcell transplantation: An active regimen for aggressive mantle-cell lymphoma. J Clin Oncol 16:3803-3809, 1998
- 18. Hay N, Sonenberg N: Upstream and downstream of mTOR. Genes Dev 18:1926-1945, 2004
- 19. Bjornsti MA, Houghton PJ: The TOR pathway: A target for cancer therapy. Nat Rev Cancer 4:335-348, 2004
- 20. Brunn GJ, Hudson CC, Sekulic A, et al: Phosphorylation of the translational repressor PHAS-I by the mammalian target of rapamycin. Science 277:99- 101, 1997
- 21. Burnett PE, Barrow RK, Cohen NA, et al: RAFT1 phosphorylation of the transla-

activity in relapsed MCL. The response rates appear similar at 250mg weekly and 25mg weekly.

This study demonstrates that agents that selectively target cellular pathways dysregulated in MCL cells can produce therapeutic benefit and this effect may not be dose dependant. Further studies of this agent in MCL and other lymphoid malignancies are therefore warranted.

> tional regulators p70 S6 kinase and 4E-BP1. Proc Natl Acad Sci U S A 95:1432- 1437, 1998

- 22. Gingras AC, Raught B, Sonenberg N: Regulation of translation initiation by FRAP/mTOR. Genes Dev 15:807-826, 2001
- 23. Gingras AC, Gygi SP, Raught B, et al: Regulation of 4E-BP1 phosphorylation: A novel two-step mechanism. Genes Dev 13:1422-1437, 1999
- 24. Schmelzle T, Hall MN: TOR, a central controller of cell growth. Cell 103:253- 262, 2000
- 25. Grolleau A, Bowman J, Pradet-Balade B, et al: Global and specific translational control by rapamycin in T cells uncovered by microarrays and proteomics. J Biol Chem 277:22175-22184, 2002
- 26. Vivanco I, Sawyers CL: The phosphatidylinositol 3-Kinase AKT pathway in human cancer. Nat Rev Cancer 2: 489-501, 2002
- 27. Huang S, Houghton PJ: Targeting mTOR signaling for cancer therapy. Curr Opin Pharmacol 3:371-377, 2003
- Dancey JE: Clinical development of mammalian target of rapamycin inhibitors. Hematol Oncol Clin North Am 16:1101-4, 2002
- 29. Mita MM, Mita A, Rowinsky EK: Mammalian target of rapamycin: A new molecular target for breast cancer. Clin Breast Cancer 4:126-137, 2003
- 30. Tolcher AW: Novel therapeutic molecular targets for prostate cancer: The mTOR signaling pathway and epidermal growth factor receptor. J Urol 171:S41- S44, 2004
- 31. Hidalgo M: New target, new drug, old paradigm. J Clin Oncol 22:2270-2272, 2004
- 32. Witzig TE, Geyer SM, Ghobrial I, Inwards DJ, Fonseca R, Kurtin P, et al. Phase II Trial of Single-Agent Temsirolimus (CCI-779) for Relapsed Mantle Cell Lymphoma. J Clin Oncol. 2005 Jun 27; [Epub .
ahead of print].
- 33. Witzig TE, Ansell SM, Geyer SM, Kurtin PJ, Rowland KM, Flynn PJ, et al. Anti-Tumor Activity of Low-Dose Single Agent CCI-779 for Relapsed Mantle Cell Lymphoma: A Phase II Trial in the North Central Cancer Treatment Group. J Clin Oncol 23(16):561s, 2005 (suppl 1).